

Table 1: Summary of nano-formulated polyphenols for improved pharmacokinetics and use in cardiovascular therapy

Polyphenol	Nano-formulation	Process	Size (nm)	Entrapment efficiency	Study type	Major results	Ref
Quercetin	Nano-quercetin	Anti-solvent precipitation	170	-	In vitro dissolution study	Enhanced quercetin dissolution in simulated intestinal fluid	18
	PLGA nanoparticle	Electrohydrodynamic atomization	384.9	-	In vitro stability and release study	Sustained release for over 59 days.	19
	SLN	Emulsification and low temperature solidification	155.3	91.1%	In situ absorption study	Enhanced intestinal absorption and oral bioavailability	20
	PLGA nanoparticle	Solvent displacement	399.67±10.86	79.84±2.5%	In vitro release and anti-oxidant study	Higher free radical scavenging activity of encapsulated flavonoid	21
	Eudragit® E and PVA nanoparticle	Nanoprecipitation	81.9±0.26	99.9±0.59%	Development and in-vitro anti-oxidant activity of nanoparticles	Greater anti-oxidant activity of nano-formulation.	22
	Mesoporous silica nanoparticle	-	100-150	89.47±1.28%	In-vitro and In-vivo models of myocardial ischemia-reperfusion injury	Mesoporous silica nanoparticles intensified the activation of JAK/STAT pathway and more effectively inhibited myocardial injury	23
	PLGA nanoparticle	Oil-in-water emulsification	165±7.5	98.15%	In-vitro model of hypoxia-reoxygenation injury	Better flavonoid delivery to cardiomyocyte upon nano-encapsulation. Increased cell	27

						viability. Reduced mitochondrial oxidative stress.	
	Superparamagnetic silica and PLGA nanoparticle	Sol-gel technique	100-200	-	Fabrication and in vitro evaluation of nano biocarrier	Controlled drug release. Reduced infarct size.	30
Curcumin	SLN	Microemulsion technique	450	70%	Formulation and characterization of nanoparticles	Prolonged drug release up to 12h. Improved physical and chemical storage stability.	40
	PBCA (poly(butyl) cyanoacrylate) nanoparticles coated with poloxamer 188	Anionic polymerization using solvent evaporation method	178	77.99%	Physicochemical characterization and stability study	Controlled drug release with higher release at acidic pH. Increased storage stability.	41
	Liposome (lecithin-SLP- white and SLP- PC70)	-	263±86	68%	In vivo oral bioavailability and anti-oxidant activity	Improved bioavailability. Enhanced anti- oxidant effect	42
	PLGA-PEG blend nanoparticles	Single-emulsion solvent evaporation	<200	70%	Oral pharmacokinetic study	Prolonged drug release. 7.4-times increase in maximum plasma concentration. 55.5-times higher bioavailability of encapsulated curcumin compared to aqueous solution	43
	PLGA-PVA nanoparticle	Emulsion-diffusion- evaporation method	264±2	76.9%	In-vivo pharmacokinetic study	9-fold increase in oral	44

						bioavailability	
	CMC (carboxymethyl chitosan) nanoparticle conjugated with myocyte specific homing peptide	-	331.2	61.24%	In-vivo and in-vitro models of cardiac hypertrophy	Targeted delivery of drug at the hypertrophic myocyte. High cardiac bioavailability of drug even at a low dose. Improved cardiac functions and reduced apoptosis of hypertrophied cardiac cells.	46
	Curcumin nanoparticles	-	300 ± 50	-	Pulmonary arterial hypertension induced cardiac remodeling	Reduced right ventricular wall thickness and tissue weight. Mitigated cardiac expression of inflammatory proteins like TNF- α and IL- 1 β . Inhibited cardiac remodeling and oxidative stress.	47
	PEG-PDLLA (poly (ethylene glycol)-b-poly (DL-lactide))	-	<100	-	In-vitro palmitate induced cardiomyocyte injury	Mitigated cardiac apoptosis and attenuated oxidative stress and lipotoxic cellular damage through the AMPK pathway	48
	PEG-PDLLA nanoparticle (Monomethoxy)	-	50	-	In-vitro cardio protection from palmitate induced	Anti-apoptotic and anti- oxidant effect	49

	poly (ethylene-glycol)-b-poly (DL-lactide))				apoptosis	reduced cardiomyocyte death. Anti-apoptotic effect mediated through inhibition of NADPH-mediated oxidative stress and ERS (endoplasmic reticulum stress)-related pathway.	
Curcumin-Nisin	PLA (poly Lactic Acid) nanoparticles	Double emulsion-diffusion-evaporation	284±17.9	-	Isoproterenol induced myocardial infarction	Reduced expression of cardiac and renal injury markers. Improved cardiac functions	50
Resveratrol-curcumin (5:1)	Pluronic® F-127 micelles	Solvent casting method	25.05	-	In-vitro doxorubicin induced cardiotoxicity	Alleviated doxorubicin induced cardiotoxicity by anti-apoptotic and anti-oxidant pathways.	45
Resveratrol-quercetin	Pluronic® F-127 micelles	Solvent casting method	22.34±0.15	-	In-vitro and in-vivo model of doxorubicin induced cardiotoxicity	Increased circulation time of enclosed molecules. Reduced oxidative stress and improved cell viability. Enhanced cardio protection.	73
	Sodium deoxycholate (SDC)- elastic	Thin lipid film method	149	97%	Formulation and characterization of dual-loaded liposome	Improved storage stability.	74

	liposome					Sustained release of drugs for 60h	
Resveratrol	PLGA nanoparticles	Emulsion evaporation	257.90±8.29nm	20%	In-vivo Isoproterenol-induced myocardial infarction	Reduced oxidative stress. Reduced inflammatory damage. Upregulated eNOS and downregulated iNOS expression. Enhanced bioactivity and cardio protection.	71
	SLN (egg yolk lecithin, glyceryl monostearate)	Emulsion-diffusion	271.13	-	In vivo pharmacokinetic and dynamic effect on animal model of doxorubicin induced cardiotoxicity	Improved solubility and sustained release of the drug. Improved cardiac functions.	72
	SLN (Stearic acid, Phospholipon® 90 G)	Solvent diffusion-solvent evaporation method	134±7.6	88.9%	Oral pharmacokinetic study	Sustained drug release up to 120h. 8-fold increase in oral bioavailability	53
	Layer-by-layer nano-formulation (polyallylamine hydrochloride and dextran sulfate)	-	215.3±2.3	-	Oral bioavailability study	1.76-fold increment in plasma drug concentration. Improved chemical stability of resveratrol in simulated gastric and intestinal fluids.	57
	SLN (N-trimethyl chitosan graft	Emulsification and ultrasonication method	258.28±18.74	95.45±2.18%	In-vitro and in-vivo drug release study	Negligible drug release in simulated	55

	palmitic acid)					gastric fluid. Sustained drug release at intestinal pH. 3.8-fold increase in oral bioavailability	
	SLN and NLC	Hot homogenization	150-250	70%	In-vitro oral drug release study	Improved storage stability. Controlled drug release at gastrointestinal pH.	54
	Carboxymethyl chitosan	Emulsion cross-linking	155.3±15.2	44.5±2.2%	In vitro anti-oxidant and in vivo bioavailability study	Enhanced anti-oxidant activity. Increased solubility and bioavailability.	58
	Galactosylated PLGA nanoparticles	Solvent diffusion method	108.4	-	Pharmacokinetic study	Enhanced oral bioavailability by 335.7%.	60
	PLGA nanoparticles	Nanoprecipitation	90.35±1.2	78.25±1.2	Oral bioavailability study	10-fold increase in bioavailability	59
Baicalin	NLC (glycerol monostearate and medium chain triglyceride)	Emulsion-evaporation and low temperature solidification	244.7	59.51±0.57%	Formulation, in-vitro drug release and in vivo pharmacokinetic study	Sustained drug release for over 48h. Significantly improved plasma drug concentration and retention time. Enhanced oral bioavailability	89
	NLC (glycerol monostearate, polyethylene glycol monostearate and oleic acid)	Emulsion-evaporation and low temperature solidification	83.9	83.5%	In-vitro and in-vivo pharmacokinetic and biodistribution evaluation	Sustained release, improved AUC and plasma retention. Increased drug accumulation at the heart. Reduced infarct size.	90

EGCG (epigallocatechin-3-gallate)	Chitosan coated NLC	Phase-inversion technique	56.5±3.2	99%	In-vitro drug release, cytotoxicity and bioactivity study on macrophages	Improved drug stability. Increased cellular drug uptake. Reduced macrophage cholesterol content and monocyte chemoattractant protein-1 (MCP-1).	94
	Gold nanoparticles	-	40±5	48%	In-vitro stability, affinity and selectivity study towards smooth muscle and endothelial cells	Selective anti-proliferative and anti-migratory effect on smooth muscle cells with no toxic effect on endothelial cells. Cellular endocytosis of EGCG via the laminin receptor	93
	Chitosan-tripolyphosphate nanoparticles	-	440±37	-	In-vivo bioavailability study	1.5-fold increase in plasma and 2.3-fold increase in intestinal (jejunum) drug concentration. Enhanced oral bioavailability	95
Icariin	Monomethyl ether polyethylene glycol (mPEG)	-	143.3	32%	In-vitro model of myocardial ischemia	Prolonged drug release for 72h. Reduced ischemic cell damage. Improved cardiomyocyte activity under glucose and	100

						oxygen deprivation. Decreased myocardial apoptosis. Enhanced icariin bioactivity.	
Puerarin	SLN (monostearin and soy lecithin)	Solvent injection method	160	-	Oral pharmacokinetic study	Increased drug plasma concentration and C _{max} with shorter T _{max} . Higher cardiac and brain drug accumulation. 1.46-times higher bioavailability.	101
	Cyclic arginyl-glycyl-aspartic acid (RGD) peptide modified PEGylated SLNs	Solvent-evaporation method	70-110	88.9±3.1%	Pharmacokinetic and dynamic study on acute myocardial ischemia in-vivo	Drug accumulation at the ischemic myocardium. Longer plasma retention time. Improved therapeutic efficacy of puerarin.	103
Puerarin prodrug-tanshinone	Co-loaded SLN	Single emulsification-solvent evaporation method	112.6±3.1	86.7±3.3%	In-vitro and in-vivo evaluation of co-loaded nanoparticle	3.4-fold increase in heart drug concentration. Increased therapeutic efficacy of enclosed drugs. Reduced infarct size with dual loaded nanoparticles compared to single drug loaded SLN.	102
Tilianin	Nano-micelles	-	70	-	In-vitro model of myocardial	Improved anti-oxidant and	105

					hypoxia/reoxygenation injury	anti-apoptotic activity. Increased cardiac protection against ischemic injury.	
Wogonin	PLGA-PVA nanoparticles	-	194.82	74.89%	In-vivo model of isoproterenol induced myocardial infarction	Significantly reduced myocardial infarct size. Enhanced cardio protection by down-regulation of inflammatory markers and upregulation of cardioprotective proteins (Nrf2 and heme oxygenase-1)	104

References:

18. Gormaz J, Quintremil S, Rodrigo R. Cardiovascular Disease: A Target for the Pharmacological Effects of Quercetin. *CTMC*. 2015;15(17):1735-1742. doi:10.2174/1568026615666150427124357
19. Murota K, Terao J. Antioxidative flavonoid quercetin: implication of its intestinal absorption and metabolism. *Archives of Biochemistry and Biophysics*. Published online 2003;6.
20. Kakran M, Sahoo NG, Li L, Judeh Z. Fabrication of quercetin nanoparticles by anti-solvent precipitation method for enhanced dissolution. *Powder Technology*. 2012;223:59-64. doi:10.1016/j.powtec.2011.08.021
21. Giannouli M, Karagkiozaki V, F.Pappa, Moutsios I, C.Gravalidis, Logothetidis S. Fabrication of quercetin-loaded PLGA nanoparticles via electrohydrodynamic atomization for cardiovascular disease. *Materials Today: Proceedings*. 2018;5(8):15998-16005. doi:10.1016/j.matpr.2018.05.044

22. Li H, Zhao X, Ma Y, Zhai G, Li L, Lou H. Enhancement of gastrointestinal absorption of quercetin by solid lipid nanoparticles. *Journal of Controlled Release*. 2009;133(3):238-244. doi:10.1016/j.jconrel.2008.10.002
23. Pool H, Quintanar D, Figueroa J de D, et al. Antioxidant Effects of Quercetin and Catechin Encapsulated into PLGA Nanoparticles. *Journal of Nanomaterials*. 2012;2012:1-12. doi:10.1155/2012/145380
27. Manukyan MC, Alvernaz CH, Poynter JA, et al. Interleukin-10 protects the ischemic heart from reperfusion injury via the STAT3 pathway. *Surgery*. 2011;150(2):231-239. doi:10.1016/j.surg.2011.05.017
30. Platel A, Carpentier R, Becart E, Mordacq G, Betbeder D, Nesslany F. Influence of the surface charge of PLGA nanoparticles on their *in vitro* genotoxicity, cytotoxicity, ROS production and endocytosis: Charge-dependent nanoparticle toxicity. *J Appl Toxicol*. 2016;36(3):434-444. doi:10.1002/jat.3247
40. Wang NP, Wang ZF, Tootle S, Philip T, Zhao ZQ. Curcumin promotes cardiac repair and ameliorates cardiac dysfunction following myocardial infarction: Curcumin and cardiac repair. *British Journal of Pharmacology*. 2012;167(7):1550-1562. doi:10.1111/j.1476-5381.2012.02109.x
41. Sunagawa Y, Funamoto M, Shimizu K, et al. Curcumin, an Inhibitor of p300-HAT Activity, Suppresses the Development of Hypertension-Induced Left Ventricular Hypertrophy with Preserved Ejection Fraction in Dahl Rats. *Nutrients*. 2021;13(8):2608. doi:10.3390/nu13082608
42. Ji X, Xiao J, Sheng X, Zhang X, Guo M. Curcumin protects against myocardial infarction-induced cardiac fibrosis via SIRT1 activation in vivo and in vitro. *DDDT*. Published online March 2016:1267. doi:10.2147/DDDT.S104925
43. Pourbagher-Shahri AM, Farkhondeh T, Ashrafizadeh M, Talebi M, Samargahndian S. Curcumin and cardiovascular diseases: Focus on cellular targets and cascades. *Biomedicine & Pharmacotherapy*. 2021;136:111214.
45. Siviero A, Gallo E, Maggini V, et al. Curcumin, a golden spice with a low bioavailability. *Journal of Herbal Medicine*. 2015;5(2):57-70. doi:10.1016/j.hermed.2015.03.001
46. Tiyaboonchai W, Tungpradit W, Plianbangchang P. Formulation and characterization of curcuminoids loaded solid lipid nanoparticles. *International Journal of Pharmaceutics*. 2007;337(1-2):299-306. doi:10.1016/j.ijpharm.2006.12.043

47. Mulik R, Mahadik K, Paradkar A. Development of curcuminoids loaded poly(butyl) cyanoacrylate nanoparticles: Physicochemical characterization and stability study. *European Journal of Pharmaceutical Sciences*. 2009;37(3-4):395-404. doi:10.1016/j.ejps.2009.03.009
48. Takahashi M, Uechi S, Takara K, Asikin Y, Wada K. Evaluation of an Oral Carrier System in Rats: Bioavailability and Antioxidant Properties of Liposome-Encapsulated Curcumin. *J Agric Food Chem*. 2009;57(19):9141-9146. doi:10.1021/jf9013923
49. Khalil NM, Nascimento TCF do, Casa DM, et al. Pharmacokinetics of curcumin-loaded PLGA and PLGA-PEG blend nanoparticles after oral administration in rats. *Colloids and Surfaces B: Biointerfaces*. 2013;101:353-360. doi:10.1016/j.colsurfb.2012.06.024
50. Shaikh J, Ankola DD, Beniwal V, Singh D, Kumar MNVR. Nanoparticle encapsulation improves oral bioavailability of curcumin by at least 9-fold when compared to curcumin administered with piperine as absorption enhancer. *European Journal of Pharmaceutical Sciences*. 2009;37(3-4):223-230. doi:10.1016/j.ejps.2009.02.019
53. Rice KM, Manne NDPK, Kolli MB, et al. Curcumin nanoparticles attenuate cardiac remodeling due to pulmonary arterial hypertension. *Artificial Cells, Nanomedicine, and Biotechnology*. 2016;44(8):1909-1916. doi:10.3109/21691401.2015.1111235
54. Zhang J, Wang Y, Bao C, et al. Curcumin-loaded PEG-PDLLA nanoparticles for attenuating palmitate-induced oxidative stress and cardiomyocyte apoptosis through AMPK pathway. *Int J Mol Med*. Published online June 5, 2019. doi:10.3892/ijmm.2019.4228
57. Burgos-Morón E, Calderón-Montaña JM, Salvador J, Robles A, López-Lázaro M. The dark side of curcumin. *Int J Cancer*. Published online 2010:NA-NA. doi:10.1002/ijc.24967
58. Walle T. Bioavailability of resveratrol: Resveratrol bioavailability. *Annals of the New York Academy of Sciences*. 2011;1215(1):9-15. doi:10.1111/j.1749-6632.2010.05842.x
59. Pandita D, Kumar S, Poonia N, Lather V. Solid lipid nanoparticles enhance oral bioavailability of resveratrol, a natural polyphenol. *Food Research International*. 2014;62:1165-1174. doi:10.1016/j.foodres.2014.05.059

60. Reis S, Neves, Lúcio, Martins, Lima. Novel resveratrol nanodelivery systems based on lipid nanoparticles to enhance its oral bioavailability. *IJN*. Published online January 2013:177. doi:10.2147/IJN.S37840
71. Tomé-Carneiro J, González M, Larrosa M, et al. Consumption of a grape extract supplement containing resveratrol decreases oxidized LDL and ApoB in patients undergoing primary prevention of cardiovascular disease: A triple-blind, 6-month follow-up, placebo-controlled, randomized trial. *Mol Nutr Food Res*. 2012;56(5):810-821. doi:10.1002/mnfr.201100673
72. Tomé-Carneiro J, González M, Larrosa M, et al. Resveratrol in primary and secondary prevention of cardiovascular disease: a dietary and clinical perspective: Resveratrol and cardiovascular disease. *Ann NY Acad Sci*. 2013;1290(1):37-51. doi:10.1111/nyas.12150
73. Goh KP, Lee HY, Lau DP, Supaat W, Chan YH, Koh AFY. Effects of Resveratrol in Patients with Type 2 Diabetes Mellitus on Skeletal Muscle SIRT1 Expression and Energy Expenditure. *International Journal of Sport Nutrition and Exercise Metabolism*. 2014;24(1):2-13. doi:10.1123/ijsnem.2013-0045
74. Chachay VS, Macdonald GA, Martin JH, et al. Resveratrol Does Not Benefit Patients With Nonalcoholic Fatty Liver Disease. *Clinical Gastroenterology and Hepatology*. 2014;12(12):2092-2103.e6. doi:10.1016/j.cgh.2014.02.024
89. Wang X, He F, Liao Y, et al. Baicalin pretreatment protects against myocardial ischemia/reperfusion injury by inhibiting mitochondrial damage-mediated apoptosis. *International Journal of Cardiology*. 2013;168(4):4343-4345. doi:10.1016/j.ijcard.2013.05.077
90. de Oliveira MR, Nabavi SF, Habtemariam S, Erdogan Orhan I, Daglia M, Nabavi SM. The effects of baicalein and baicalin on mitochondrial function and dynamics: A review. *Pharmacological Research*. 2015;100:296-308. doi:10.1016/j.phrs.2015.08.021
93. Taiming L, Xuehua J. Investigation of the absorption mechanisms of baicalin and baicalein in rats. *Journal of Pharmaceutical Sciences*. 2006;95(6):1326-1333. doi:10.1002/jps.20593
94. Xing J, Chen X, Zhong D. Absorption and enterohepatic circulation of baicalin in rats. *Life Sciences*. 2005;78(2):140-146. doi:10.1016/j.lfs.2005.04.072

95. Luan J, Zheng F, Yang X, Yu A, Zhai G. Nanostructured lipid carriers for oral delivery of baicalin: In vitro and in vivo evaluation. *Colloids and Surfaces A: Physicochemical and Engineering Aspects*. 2015;466:154-159. doi:10.1016/j.colsurfa.2014.11.015
100. Zhang J, Nie S, Wang S. Nanoencapsulation Enhances Epigallocatechin-3-gallate Stability and Its Antiatherogenic Bioactivities in Macrophages. *J Agric Food Chem*. 2013;61(38):9200-9209. doi:10.1021/jf4023004
101. Dube A, Nicolazzo JA, Larson I. Chitosan nanoparticles enhance the plasma exposure of (–)-epigallocatechin gallate in mice through an enhancement in intestinal stability. *European Journal of Pharmaceutical Sciences*. 2011;44(3):422-426. doi:10.1016/j.ejps.2011.09.004
102. Ke Z, Liu J, Xu P, Gao A, Wang L, Ji L. The Cardioprotective Effect of Icariin on Ischemia-Reperfusion Injury in Isolated Rat Heart: Potential Involvement of the PI3K-Akt Signaling Pathway. *Cardiovasc Ther*. 2015;33(3):134-140. doi:10.1111/1755-5922.12121
103. Scicchitano M, Carresi C, Nucera S, et al. Icariin Protects H9c2 Rat Cardiomyoblasts from Doxorubicin-Induced Cardiotoxicity: Role of Caveolin-1 Upregulation and Enhanced Autophagic Response. *Nutrients*. 2021;13(11):4070. doi:10.3390/nu13114070
104. Fang J, Zhang Y. Icariin, an Anti-atherosclerotic Drug from Chinese Medicinal Herb Horny Goat Weed. *Front Pharmacol*. 2017;8:734. doi:10.3389/fphar.2017.00734
105. Xu S, Yu J, Zhan J, Yang L, Guo L, Xu Y. Pharmacokinetics, Tissue Distribution, and Metabolism Study of Icariin in Rat. *BioMed Research International*. 2017;2017:1-17. doi:10.1155/2017/4684962