SUPPLEMENTAL MATERIALS

Renal single-nuclear transcriptomics identifies novel therapeutic targets in a preclinical model of chronic kidney disease.

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Supplemental Tables

Supplemental Table 1a. Composition of the high cholesterol/high carbohydrate diet (KT324). For a complete list of ingredients, please see Supplementary Table 1b.

Ingredient	%
5B4L LabDiet version	79.2223%
Hydrogenated Soy Oil	8.4000%
High Fructose Corn Syrup - 55	5.0000%
Hydrogenated Coconut Oil	4.7000%
Cholesterol	2.0000%
Sodium Cholate	0.6777%
Energy (kcal/g)	%
Protein	16.3
Fat (ether extract)	42.9
Carbohydrates	40.8

Supplemental Table 1b. Full list of ingredients of KT324 diet.

Complete Ingredients List	Measurements (%, ppm,IU/g,mcg/kg)
Arginine	0.92%
Histidine	0.47%
Isoleucine	0.95%
Leucine	1.44%
Lysine	1.16%
Methionine	0.36%
Cystine	0.17%
Phenylalanine	0.86%
Tyrosine	0.71%
Threonine	0.67%
Tryptophan	0.22%
Valine	1.09%
Alanine	0.64%
Aspartic Acid	1.54%
Glutamic Acid	3.04%
Glycine	0.56%

Proline	1.09%
Serine	0.81%
Taurine	0.00%
Cholesterol	20,000 ppm
Linoleic Acid	1.45%
Linolenic Acid	0.13%
Arachidonic Acid	0.00%
Omega-3 Fatty Acid	0.13%
Total Saturated Fatty Acids	10.69%
Total Monounsaturated Fatty Acids	4.98%
Polyunsaturated Fatty Acids	1.33%
Neutral Detergent Fiber	11.90%
Acid Detergent Fiber	6.20%
Starch	28.35%
Glucose	2.29%
Fructose	18.64%
Sucrose	1.67%
Lactose	0.01%
Ash	5.90%
Calcium	1.03%
Phosphorus	0.81%
Phosphorus (available)	0.65%
Potassium	0.83%
Magnesium	0.21%
Sulfur	0.13%
Sodium	0.25%
Chloride	0.38%
Fluorine	38.70%
Iron	288 ppm
Zinc	222 ppm
Manganese	40 ppm
Copper	6 ppm
Cobalt	0.30 ppm
lodine	1.14 ppm
Chromium	2.17 ppm
Selenium	0.36 ppm
Carotene	0.1 ppm
Vitamin A	6 IU/g
Vitamin D-3	1.1 IU/g
Vitamin E	110 IU/kg
Vitamin K (as menadione)	2.9 ppm
Thiamin Hydrochloride	10 ppm
Riboflavin	13.9 ppm
Niacin	64 ppm
Pantothenic Acid	40 ppm
Folic Acid	4.0 ppm
Pyridoxine	6.41 ppm
Biotin	0.4 ppm
Vitamin B-12.	78 mcg/kg
vicaniii D 12,	10 mog/ng

Supplemental Table 2. Composition of the Normal diet (ProBuild Grower and CU Sinclair S-9).

Ingredient-ProBuild Grower	%	Ingredient-CU Sinclair S-9	%
Crude Protein (Min)	18.00%	Crude Protein (Min)	19%
Lysine (Min)	1.15%	Lysine (Min)	0.90%
Crude Fat (Min)	2.00%	Crude Fat (Min)	3.75%
Crude Fiber (Max)	5.00%	Crude Fiber (Max)	3.90%
Calcium (Ca) (Min)	0.32%	Calcium (Ca) (Min)	0.55%
Calcium (Ca) (Max)	0.82%	Calcium (Ca) (Max)	1.05%
Phosphorus (P) (Min)	0.50%	Phosphorus (P) (Min)	0.65%
Salt (NaCl) (Min)	0.23%	Salt(NaCl) (Min)	0.05%
Salt (NaCl) (Max)	0.73%	Salt(NaCl) (Max)	0.55%
Selenium (Se) (Min)	0.30 ppm	Selenium (Se) (Min)	0.30 ppm
Zinc (Zn) (Min)	100.00 ppm	Zinc (Zn) (Min)	125.0 ppm
Phytase (A.Oryzae) (Min)	454 FYT/LB		

Supplemental Table 3: Alignment summary showing estimated and final number of cells in Normal and CKD kidneys (52,213 cells, n=3/group) that were subjected to quality control analysis. Normal controls: D009, D010, D011. CKD: D002, D004, D005.

	D002	D004	D005	D009	D010	D011
Estimated Number of Cells	6,771	10,035	8,912	8,707	11,068	6,367
Mean Reads per Cell	81,648	46,528	56,862	61,795	41,505	73,941
Median Genes per Cell	2,496	2,318	2,216	1,204	2,078	2,052
Number of Reads	552,841,532	466,911,820	506,756,352	538,051,075	459,378,501	470,783,915
Valid Barcodes	97.60%	97.40%	97.30%	96.60%	96.60%	96.90%
Sequencing Saturation	82.60%	73.60%	75.90%	85.90%	69.10%	80.90%
Q30 Bases in Barcode	93.60%	93.60%	93.60%	93.50%	94.20%	94.20%
Q30 Bases in RNA Read	90.50%	90.60%	91.20%	90.80%	91.10%	92.10%
Q30 Bases in UMI	93.40%	93.30%	93.30%	93.10%	93.00%	93.00%
Reads Mapped to Genome	94.80%	95.10%	95.10%	93.90%	95.20%	95.20%
Reads Mapped Confidently to Genome	91.20%	91.70%	91.70%	88.80%	92.00%	91.90%
Reads Mapped Confidently to Intergenic Regions	12.40%	12.20%	13.40%	12.50%	12.90%	12.60%
Reads Mapped Confidently to Intronic Regions	55.30%	58.90%	60.20%	53.50%	62.90%	60.90%
Reads Mapped Confidently to Exonic Regions	23.40%	20.60%	18.10%	22.90%	16.20%	18.40%
Reads Mapped Confidently to Transcriptome	64.10%	61.40%	54.30%	67.90%	50.90%	61.40%
Reads Mapped Antisense to Gene	14.40%	17.90%	23.70%	8.20%	28.00%	17.70%
Fraction Reads in Cells	74.00%	82.20%	81.10%	52.70%	82.50%	67.70%
Total Genes Detected	29,683	29,745	29,786	29,558	29,656	29,242
Median UMI Counts per Cell	5,715	5,089	4,854	1,916	4,372	4,415
After cell bender	6,490	9,895	8,855	7,826	10,822	6,027

Supplemental Table 4: Alignment summary showing the final number of cells after quality control of Normal and CKD kidneys (39,935 cells, n=3/group). Normal controls: D009, D010, D011. CKD: D002, D004, D005.

Samples	Total	Mito	Gene	UMI	Doublet	Failed	Passed
D002	6,490	21	19	139	665	812	5,678
D004	9,895	103	99	238	1,409	1,688	8,207
D005	8,855	157	188	328	1,228	1,583	7,272
D009	7,826	30	347	1,841	952	2,798	5,028
D010	10,822	102	163	454	1,663	2,136	8,686
D011	6,072	73	97	409	586	1,008	5,064

Mito: mitochondria

UMI: unique molecular identifier

Supplemental figure legends

<u>Supplemental Figure 1</u>: Single nuclei RNA-seq (snRNA-seq) workflow of Normal and CKD kidneys (n=3/group). QC: quality control; UMI: unique molecular identifier.

<u>Supplemental Figure 2</u>: snRNA-seq quality control showing analysis summary showing the percentage (%) of mitochondrial content in cells (density plots) of Normal and CKD kidneys (n=3/group). Normal controls: D009, D010, D011. CKD: D002, D004, D005.

<u>Supplemental Figure 3</u>: snRNA-seq quality control analysis summary showing the mitochondrial content detected in each cell (box plots) of Normal and CKD kidneys (n=3/group). Normal controls: D009, D010, D011. CKD: D002, D004, D005.

<u>Supplemental Figure 4</u>: snRNA-seq quality control showing the number of genes detected in each cell (density plots) of Normal and CKD kidneys (n=3/group). Normal controls: D009, D010, D011. CKD: D002, D004, D005.

<u>Supplemental Figure 5</u>: snRNA-seq quality control showing the number of genes detected in each cell (violin plots) of Normal and CKD kidneys (n=3/group). Normal controls: D009, D010, D011. CKD: D002, D004, D005.

<u>Supplemental Figure 6</u>: snRNA-seq quality control showing analysis summary showing the number of unique molecular identifiers detected in each cell (density plots) of Normal and CKD kidneys (n=3/group). Normal controls: D009, D010, D011. CKD: D002, D004, D005.

<u>Supplemental Figure 7</u>: snRNA-seq quality control showing the number of unique molecular identifiers detected in each cell (violin plots) of Normal and CKD kidneys (n=3/group). Normal controls: D009, D010, D011. CKD: D002, D004, D005.

<u>Supplemental Figure 8</u>: snRNA-seq quality control showing the estimated doublet score (scDBIFinder) detected in each cell (density plots) of Normal and CKD kidneys (n=3/group). Normal controls: D009, D010, D011. CKD: D002, D004, D005.

<u>Supplemental Figure 9</u>: snRNA-seq quality control showing the percentage of mitochondrial content vs. the number of genes detected per cell (scattered plots) of Normal and CKD kidneys (n=3/group). Normal controls: D009, D010, D011. CKD: D002, D004, D005.

<u>Supplemental Figure 10</u>: snRNA-seq quality control showing the correlation (Spearman) of genes detected vs. unique molecular identifiers (scattered plots) of Normal and CKD kidneys (n=3/group). Normal controls: D009, D010, D011. CKD: D002, D004, D005.

Supplemental Figure 11: UMAP visualization of the integrated dataset showing 39,935 nuclei of all kidneys (n=3/group). Normal controls: D009, D010, D011. CKD: D002, D004, D005.

Supplemental Figure 12: UMAP visualization of integrated dataset showing 39,935 nuclei and 30 cell clusters in the kidneys of Normal and CKD pigs (n=3/group).

<u>Supplemental Figure 13</u>: Box plot showing mitochondrial content (%), unique molecular identifier (UMI) per cell, number of genes detected per cell, and estimated doublet content in each cluster (n=3/group).

<u>Supplemental Figure 14</u>: Bar graph visualization of integrated datasets showing the distribution of cells in each cluster (n=3/group). Normal controls: D009, D010, D011. CKD: D002, D004, D005.

Supplemental Figure 15: UMAP visualization of integrated dataset showing 39,935 nuclei and 18 clusters using specific renal cell markers in Normal and CKD pigs (n=3/group).

Supplemental Figure 16: Marker gene expression profile of proximal tubular cells (UMAP) in Normal and CKD pigs (n=3/group).

Supplemental Figure 17: Marker gene expression profile of endothelial cells (UMAP) in Normal and CKD pigs (n=3/group).

<u>Supplemental Figure 18</u>: Marker gene expression profile of connecting tubular cells (UMAP) in Normal and CKD pigs (n=3/group).

<u>Supplemental Figure 19</u>: Marker gene expression profile of loop of Henle cells (UMAP) in Normal and CKD pigs (n=3/group).

Supplemental Figure 20: Marker gene expression profile of α-intercalated cells (UMAP) in Normal and CKD pigs (n=3/group).

<u>Supplemental Figure 21</u>: Marker gene expression profile of principal cells (UMAP) in Normal and CKD pigs (n=3/group).

Supplemental Figure 22: Marker gene expression profile of distal tubular cells (UMAP) in Normal and CKD pigs (n=3/group).

Supplemental Figure 23: Marker gene expression profile of podocytes (UMAP) in Normal and CKD pigs (n=3/group).

Supplemental Figure 24: Marker gene expression profile of renal interstitial cells (UMAP) in Normal and CKD pigs (n=3/group).

Supplemental Figure 25: Marker gene expression profile of β-intercalated cells (UMAP) in Normal and CKD pigs (n=3/group).

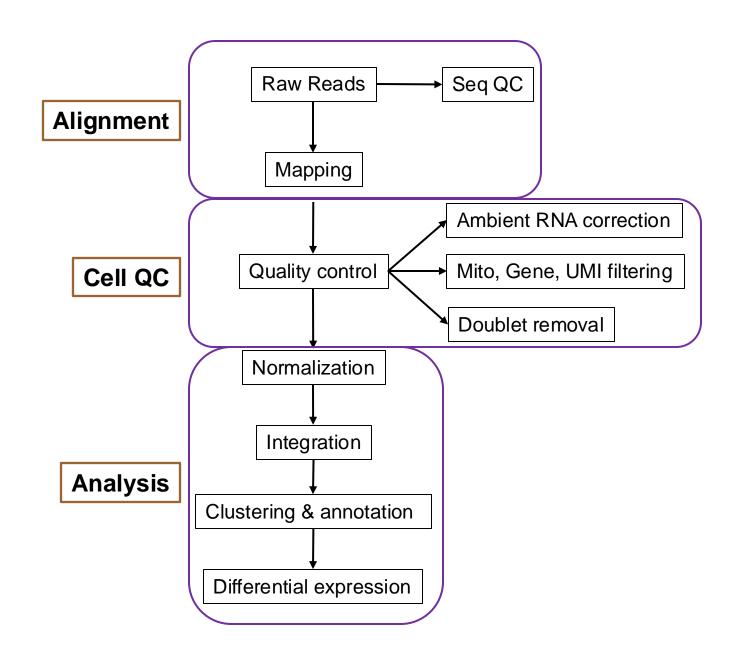
Supplemental Figure 26: Marker gene expression profile of immune cells (UMAP) in Normal and CKD pigs (n=3/group).

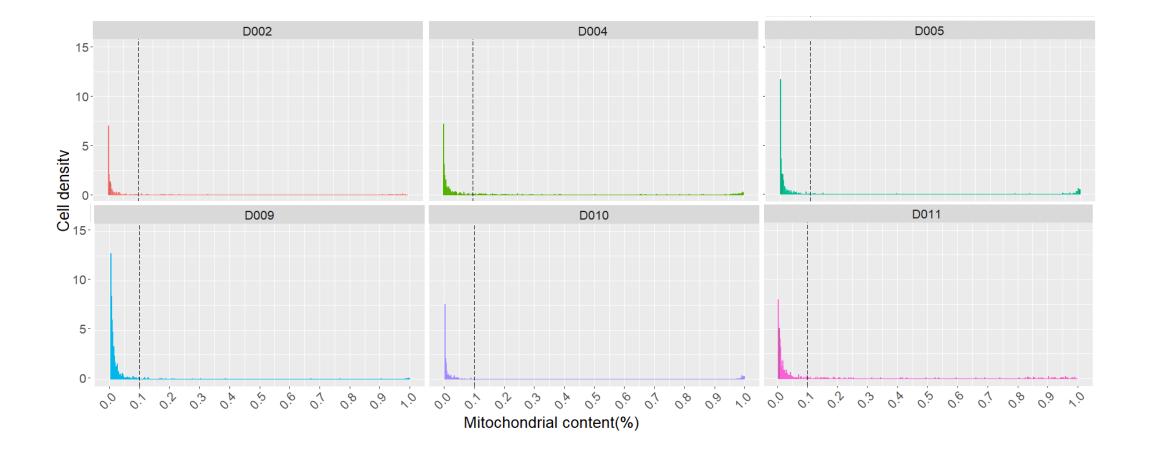
Supplemental Figure 27: Marker gene expression profile of mesangial cells (UMAP) in Normal and CKD pigs (n=3/group).

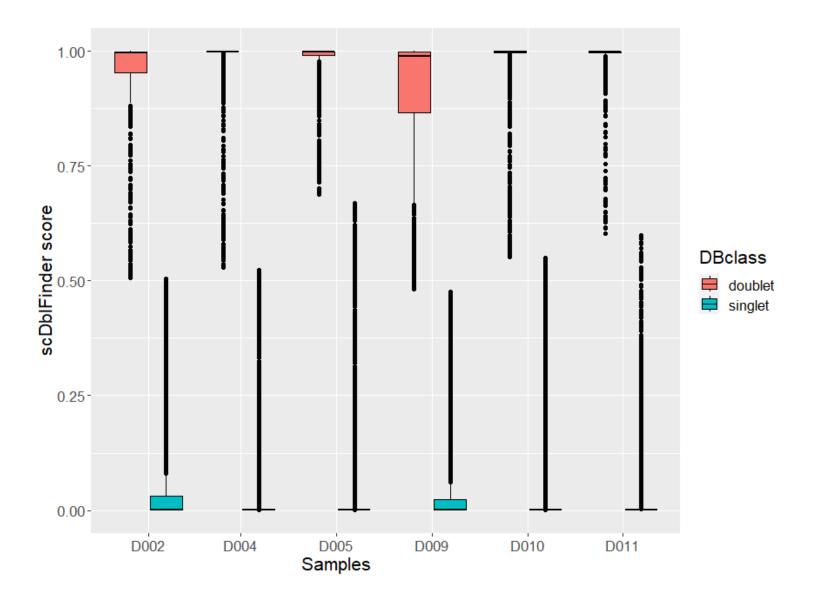
Supplemental Figure 28: Marker gene expression profile of unknown cells (UMAP, unknown 1) in Normal and CKD pigs (n=3/group).

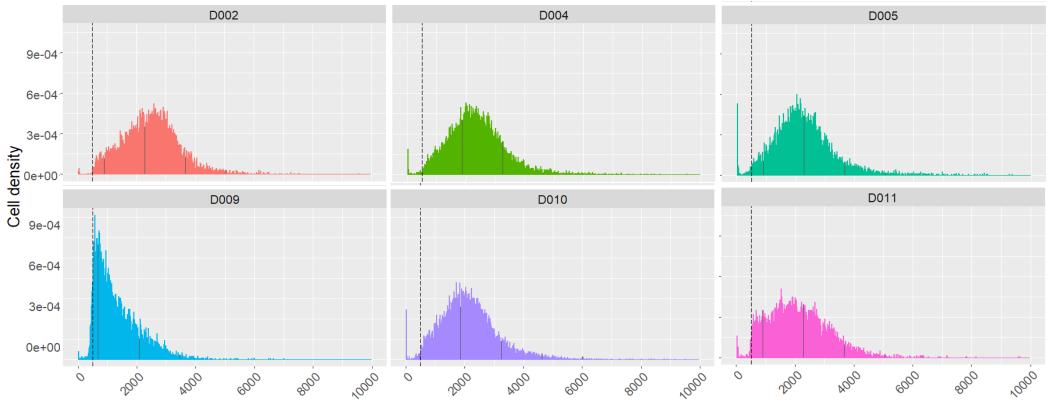
Supplemental Figure 29: Marker gene expression profile of unknown cells (UMAP, unknown 2) in Normal and CKD pigs (n=3/group).

Supplemental Figure 30: Marker gene expression profile of unknown cells (UMAP, unknown 3) in Normal and CKD pigs (n=3/group).

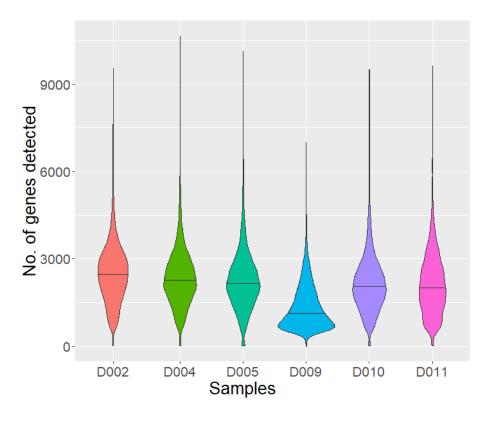


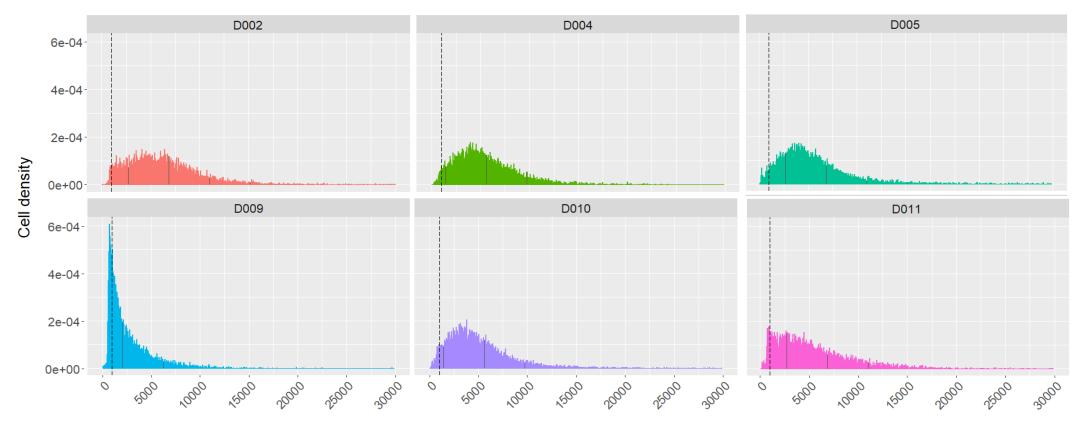




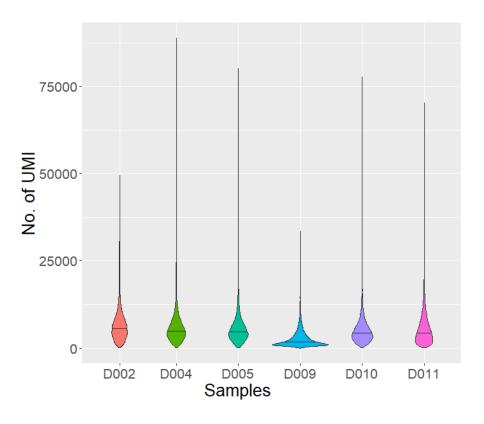


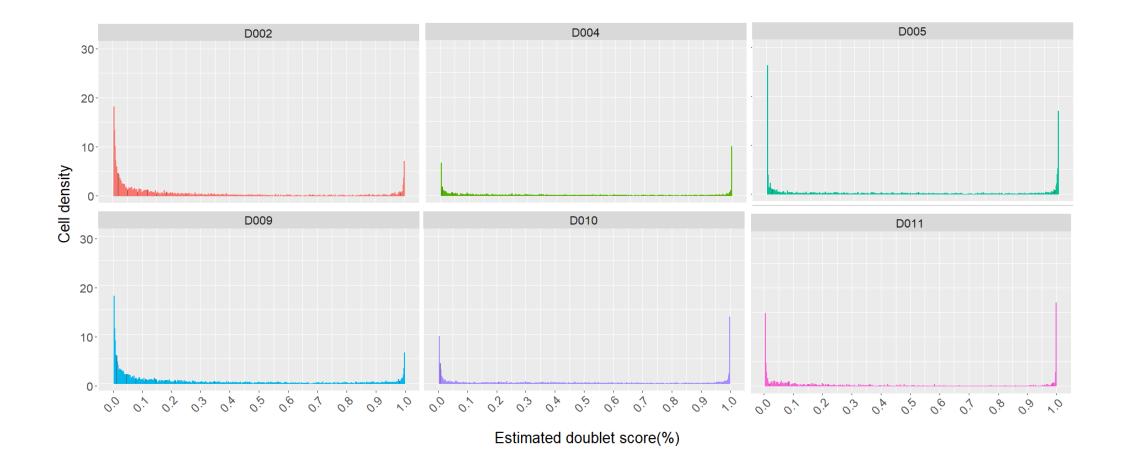
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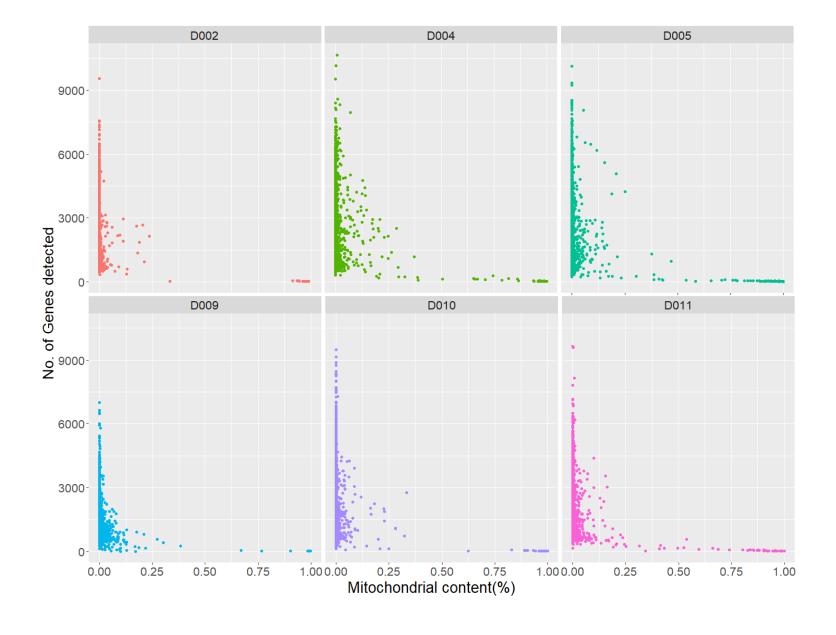


No. of unique molecular identifiers

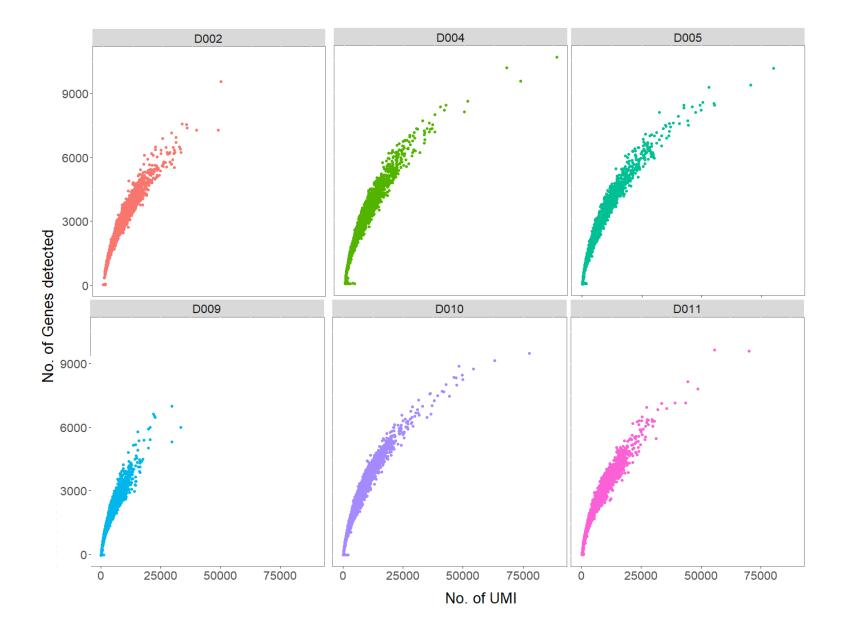




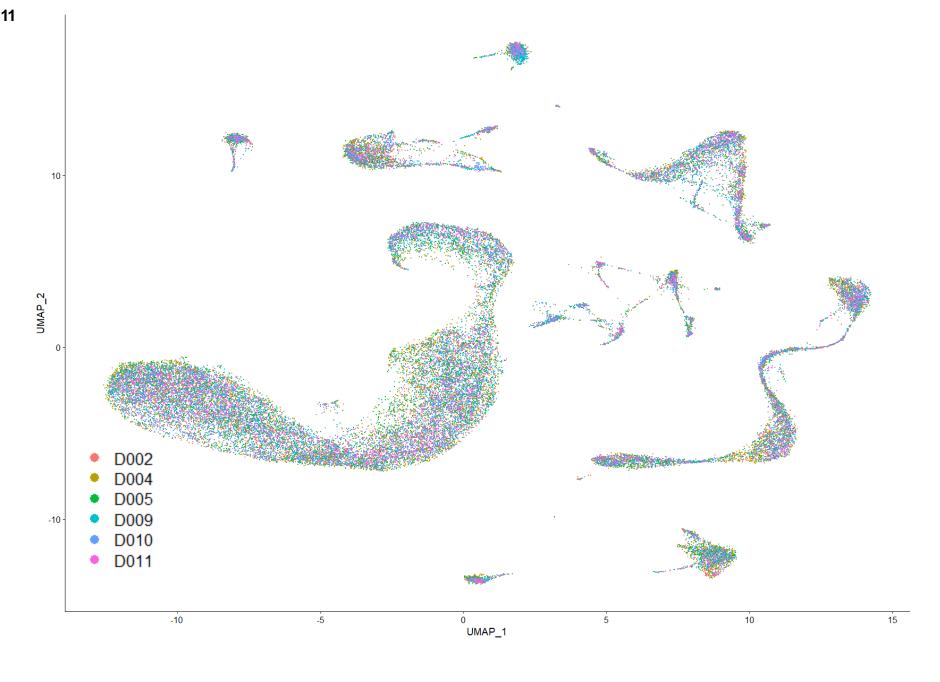
Supplemental figure 9



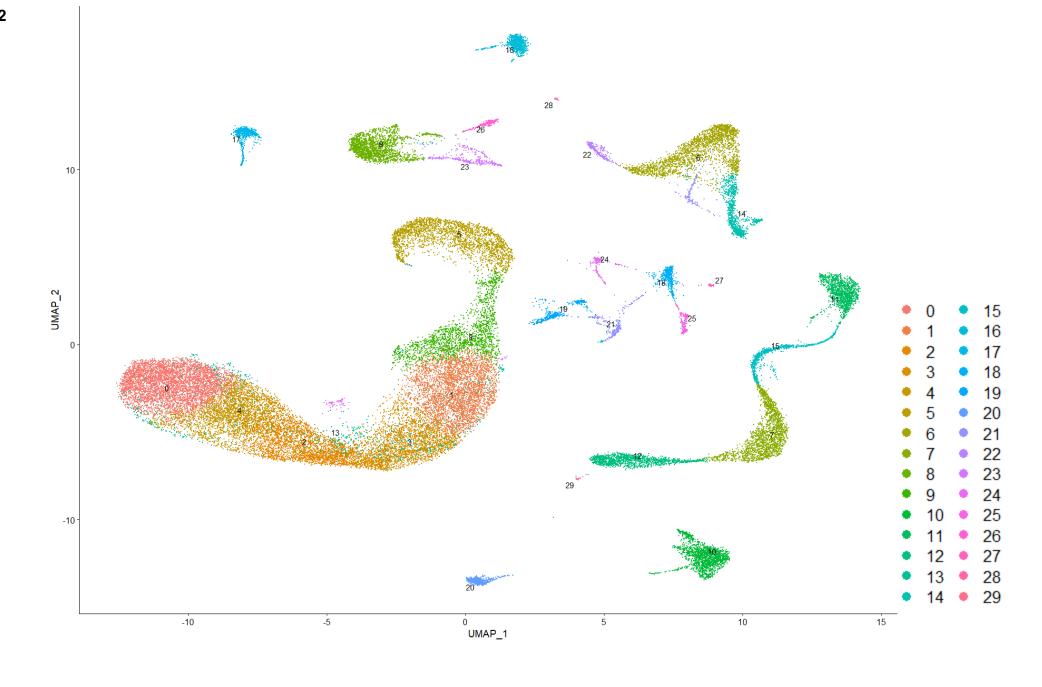
Supplemental figure 10



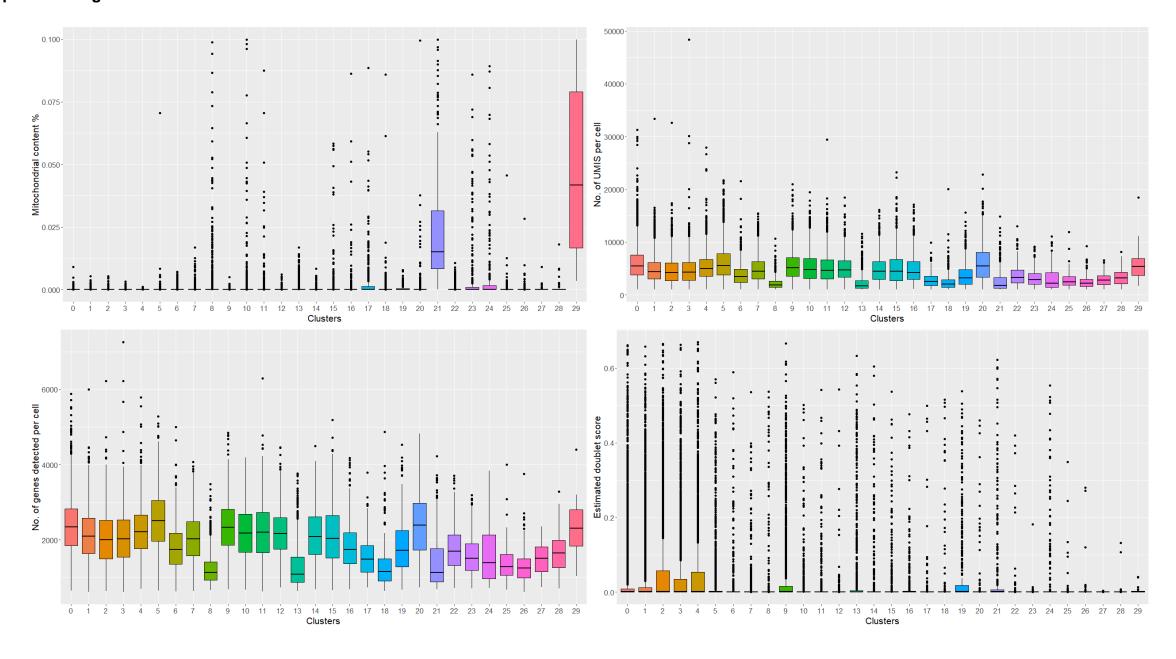
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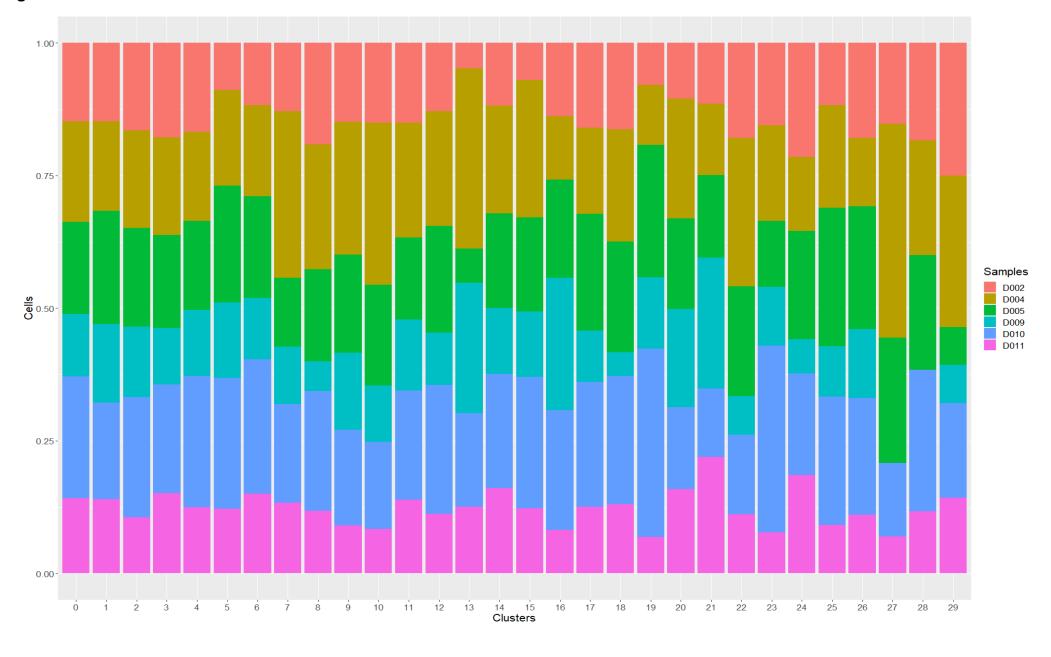
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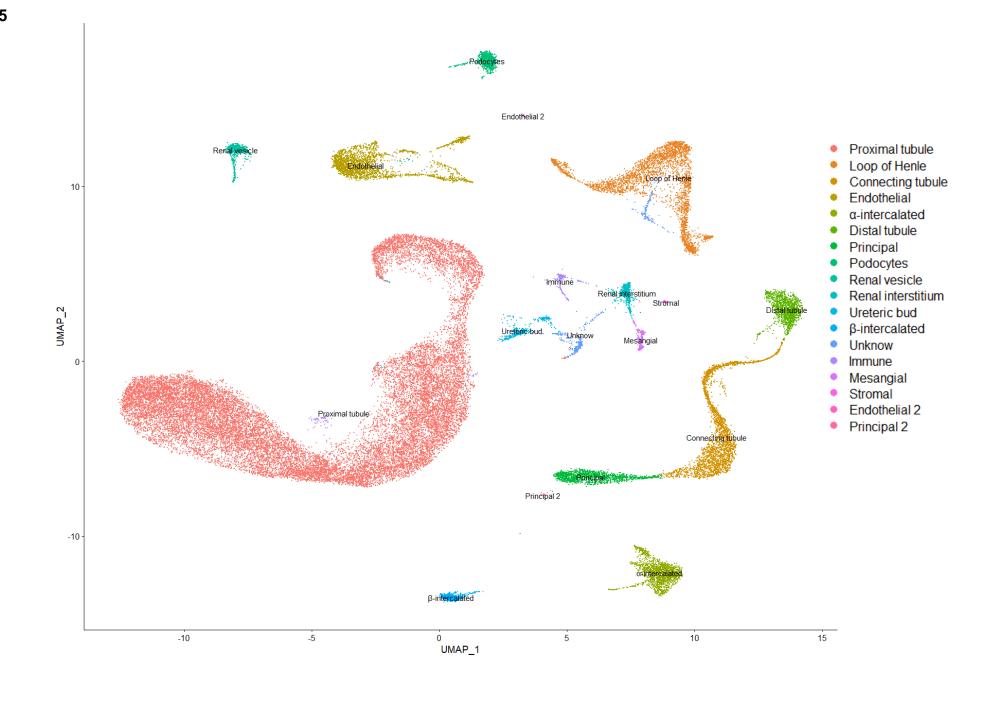
Supplemental figure 13

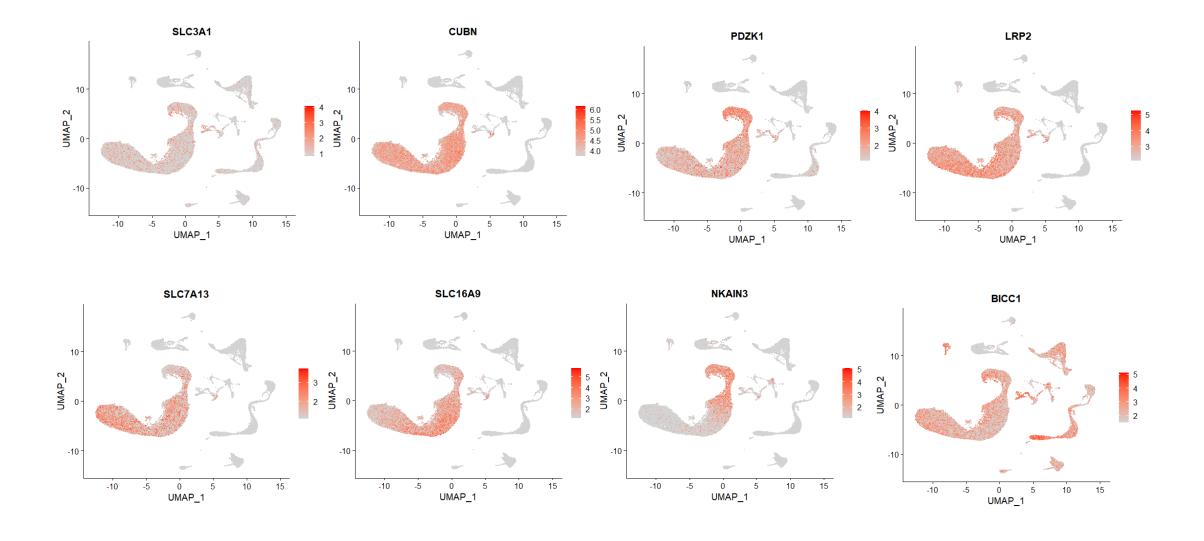


Supplemental figure 14



Supplemental figure 15





Supplemental figure 17

