Supplemental Digital Content 1 Appendix to Coriander: Overview of Potential Health Benefits Keith Singletary PhD

Antioxidant actions

The essential oil, and lipophilic and hydrophilic extracts of coriander exhibit antioxidant activity *in vitro* (1-10). For example, the radical scavenging activity of coriander seed oil was superior to that of black cumin and niger seed oils, suggesting that polar lipids may be important contributors to antioxidation (11). In another report, cilantro essential oil had one of the strongest protective effects against lipid peroxidation compared to four other essential oils and two synthetic antioxidants (12). One study reported that hydrophilic extracts of coriander leaves showed stronger antioxidant activity than that of the seeds (2). In two human cell-based assays, hydrophilic extracts of coriander seeds and leaves protected lymphocytes and keratinocytes from oxidative stress (13,14). This effect of the extracts was due in part to increasing activities of oxidative defense enzymes and levels of glutathione. Evidence for linalool's effectiveness in suppressing oxidative stress is limited and potency varies according to the assay used (15-17).

Antimicrobial actions

Essential oils of aromatic plants have been long considered important sources of antimicrobial agents (18,19). Although reportedly not as potent as those from other spices (20) oils and extracts from *Coriandrum sativum* have antibacterial activity of varying potency depending on the fraction evaluated. In general the essential oils from both the seeds and leaves (cilantro) show some of the strongest action against bacterial strains. The essential oils can inhibit both gram (+) and gram (-) bacteria and specific human pathogens including *Bacillus* species, *Salmonella* species, *Listeria monocytogenes*, *Escherichia coli*, *Staphylococcus* species, and *Streptococcus pyogenes*, although there is variability in specificity of inhibition and potency of individual coriander constituents. The inhibitory potency of coriander against the pathogen Pseudomonas aeruginosa is inconsistent (21-28). One report concluded that cilantro essential oil exerted a much more potent inhibitory effect against gram (+) bacteria, compared to that of the essential oil of seeds (21). On the other hand gram (-) were reported to be more sensitive than gram (+) bacteria to growth inhibition by the coriander seed oil (29) an effect not consistently observed (22). The profiles of inhibitory chemicals in coriander samples vary considerably, complicating the identification of the most potent constituents. For example, the essential oil of seeds was reported to consist primarily of linalool (65-90%) with lesser amounts of α -pinene, camphor and other volatiles. Yet, the essential oil of cilantro leaf in one study was reported to be composed of 26% linalool, 28% decenals and 8% 2-decen-1-ol (21), and, in another study to have 77% aliphatic alcohols and 12% aliphatic aldehydes (30). Linalool was reported to be effective against 8 periodontopathic and cariogenic bacterial strains (28,31), while a coriander essential oil fraction rich in α -pinene was more potent than one rich in linalool against the pathogens Pseudomonas, Salmonella, Listeria and Staphylococcus (21). Linalool has little antimicrobial action against *P. aeruginosa* (32). Aliphatic alcohols and aldehydes in essential oils also have antibacterial actions (33,34). For example, (2E)-dodecenal and (2E)-undecenal from the volatile oil of coriander leaves were most bactericidal against Salmonella among those similar compounds tested (35). Taken together, these studies indicate that several oil constituents are likely to contribute to inhibition of bacterial growth. Coriander essential oil has been reported to synergize with antibiotics such as ciprofloxacin and tetracyclin against Acinetobacter baumanii (36). The essential oil is antibacterial by compromising cell permeability which degrades general cell functions and metabolism (26). Essential oils have played a historical role in treating respiratory tract infections because of their secretolytic actions

(25). Thus, evaluating the impact of coriander essential oil in appropriate experimental models on respiratory tract infections via inhalation or oral delivery (25) would be an additional avenue for future research. In contrast to essential oils, aqueous and alcoholic extracts have much less antibacterial efficacy against a variety of pathogenic bacterial isolates (37-42).

Essential oils of coriander seeds, leaves and stems demonstrate antifungal efficacy *in vitro*. Organisms inhibited include *Aspergillus*, *Rhodotorula*, *Rhizopus*, *Geotrichum*, *Saccharomyces* and *Candida* (21,23,25,28,30,43-46). In one report, coriander oil showed the highest level of overall fungal inhibition compared to the oils of other spices (21). Coriander seed oils contained predominantly linalool (64-90%), whereas leaf essential oil consisted of less linalool (26%) and more alcohols and aldehydes (20%). Differences in oil composition have been responsible for some disparities among coriander species in inhibiting *Aspergillus niger* and *Candida tropicalis* (25,30). Some antifungal constituents identified by bioactivity-guided fractionation include linalool, α -pinene, hexen-1-ol, and cyclodexane (21,45,46). Although the fungus *Kluveromyces* was inhibited by a coriander essential oil, it was not by an equivalent amount of linalool (28). Thus the antifungal capacity of this oil depends on the organism and the composition of the oil investigated. Antifungal mechanisms of the oil include damage to the cytoplasmic membrane leading to an impairment of all cellular functions, inhibition of germ tube formation, and interference with the morphological switch and biofilm formation (43,45,46).

There is very limited information about the inhibitory activity of coriander toward viruses and parasites (19,47,48).

Taken together these results suggest that coriander essential oil has substantial efficacy against a broad range of bacteria and fungi. Substantiation of the antimicrobial actions observed in cell culture studies needs to be rigorously confirmed in animal models of infection, and in appropriate human interventions. Coriander has been reported to suppress growth of food spoilage organisms and human pathogens in foods (26,43,49-58).

Neurological actions

Linalool affects anxiety, analgesia and cognition in animal models when administered by injection. Both antidepressant and genotoxic actions of linalool treatment (10,50,100 or 200 mg/kg; i.p.) were evaluated in mice (59). Compared to controls, linalool showed antidepressant-like activity in the tail suspension test at doses of 100 and 200 mg/kg. Furthermore, linalool did not show any genotoxic effects as measured *in vitro* using samples of peripheral blood and brain tissue. In another study in mice (60), compared to controls, linalool significantly improved performance in the forced swim (FST), rotarod and traction tests, and the open field (OFT) and exploratory cylinder (ECT) tests of anxiolytic behavior. However, the dose-responses of linalool treatments (54.8, 100 and 173.2 mg/kg, i.p.) were not consistent among the tests, and the 54.8 mg/kg dose was ineffective in all tests. In rats, linalool dosing (125 mg/kg, i.p.) showed anxiolytic actions as measured by the elevated plus maze (EPM) test (61). In a recent report there is evidence of gender-specific effects when linalool was administered to rats at a lower dose (3 mg/kg, i.p.) for 7 days prior to evaluation in the OFT and EPM (62). Gender did not affect linalool's influence on anxiety-related behavior in the OFT. In contrast, in the EPM, linalool treatment attenuated anxiety-related behavior of males but not females. The basis for this gender-based difference was not characterized.

Inhalation of linalool and calming behavior also have been examined. Linalool treatment of mice (inhalation chamber saturated with linalool at 1% or 3%) for 60 min (63) produced sedation without significantly impairing motor control (63,64). In one report inhalation of linalool (27mg vapor) by female and male mice for 30, 60 or 90 min resulted in plasma concentrations of linalool of 1.0, 2.7 and 3.0 ng/ml, respectively, which correlated with the efficacy of sedation of each dose (65). Another inhalation study (66) using additional testing models characterized that linalool treatment (1% or 3% vapors) resulted in anxiolytic responses, increased social interaction, and decreased aggressive behavior, compared to controls, although the effective doses differed among the tests. However, a concern was that memory acquisition was impaired at the higher dose of linalool. Impaired memory acquisition also was reported (67) for rats injected with linalool (50 or 100 mg/kg, i.p.). The authors suggested that the impairment was likely due to antagonism by linalool of the NMDA glutamatergic receptor. This memory impairment effect of linalool needs to be further characterized to determine how the method of administration, dose and duration of exposure, and circulating plasma levels relate to this adverse response.

Linalool has antinociceptive actions in a variety of experimental pain models. For example, linalool was administered to mice (25 to 100 mg/kg, s.c.) 30 min prior to evaluating behavior in both chemically-induced writhing and heat-induced pain tests, which model inflammatory pain and supraspinal analgesia, respectively (68). Compared to controls, the 25 to 75 mg/kg doses inhibited the chemically-induced pain response and only the 100 mg/kg dose suppressed heat-induced pain. Similar results were observed in subsequent evaluation of linalool in formalin-induced and heat-induced pain models (69,70). Improving the delivery and efficacy of linalool *in vivo* also was studied in two reports by the same group. Mice were treated with linalool (20 mg/kg and 40 mg/kg, p.o.) prior to administering chemically-induced and heatinduced pain tests (71). Linalool also was administered (25 mg/kg, p.o.) to mice in a model of chronic non-inflammatory muscle pain that mimics human fibromyalgia (72). In both studies the analgesic efficacy of linalool alone was compared to that of β-cyclodextrin-complexed linalool, with β -cyclodextrin complexing being a means to improve the circulating half-life and the pharmacological properties of the lipophilic linalool. In both reports linalool demonstrated significant anti-hyperalgesic properties, compared to controls, and the β -cyclodextrin complex significantly improved linalool's antinociceptive effect.

The mechanisms identified for linalool's actions on the nervous system are diverse. For example, in one injection study, the antidepressant-like activity of linalool was due to interactions with the monoaminergic system (73). Using freshly isolated preparations of hemidiaphragm muscles of mice (74), it was reported that exposure to linalool reduced acetylcholine release and changed nicotinic receptor-ion channel kinetics at the neuromuscular junction. In an inhalation study, the effects of linalool treatment on the plasma levels of ACTH, catecholamine and gonadotrophin were determined in ether-sedated, menopausal rats (75). Ether-sedation produced an increase in plasma ACTH levels, which were decreased in rats inhaling linalool, and linalool inhalation restored catecholamine levels to near normal after decreases due to ether sedation. Ether sedation also elevated luteinizing hormone levels, which were lowered for rats inhaling linalool. The authors suggested that linalool inhalation may relieve the tension of menopausal disorders. Two studies characterized the linalool inhalationinduced changes in gene expression profiles in rats. In one study rats were exposed to R-(-)linalool during 2 hr of restraint stress (76). Restraint stress increased plasma ACTH and corticosterone levels, which were suppressed in those rats also inhaling linalool. Similarly, the stress-associated changes in numbers of circulating neutrophils and leukocytes were returned to normal for the linalool-inhaling rats. Holistic changes in whole blood mRNA expression measured by microarray showed that linalool inhalation repressed restraint-induced changes in the expression levels of 109 genes and enhanced the changes in 6 other genes. The authors

suggested that the changes in stress-induced gene expression profiles by linalool could be due to changes in cell subsets in the whole blood and/or to effects on transcription regulation. In a subsequent study by the same authors using the same experimental model (77), changes in the expression of over 600 genes in the hypothalamus were associated with R-(-)-linalool inhalation. Linalool inhalation enhanced expression of genes related to neuron differentiation under stress conditions, specifically those regulating hypothalamic neural conductivity and synapse formation. There also was up-regulation of genes involved in cellular defense responses and those involved in the TGF- β signaling pathway. The authors suggested that further characterization of these changes in gene expression is warranted to better understand the influence of linalool inhalation on stress responses, and, in light of the role of the hypothalamus in regulating satiety, on feeding behavior (77,78).

Linalool was examined in injection models of chemical-, thermal- or mechanicallyinduced pain for any antinociceptive efficacy (79-87). Linalool treatment effectively inhibited inflammation and neuropathic pain in these reports. Several neural mechanisms of linalool action were identified in these *in vivo* studies and include activation of the opoidergic and cholinergic systems (68,69,71,79,80-84,87), modulation of A₁ and A_{2A} receptor activities (70), increasing Fos protein expression (72), decreasing extracellular signal-related protein kinase (ERK) activation (85), and inhibition of pro-inflammatory cytokines (82,86).

Antidiabetic actions

Linalool treatment (25 mg/kg, p.o., for 45 days) was evaluated in streptozotocin (STZ)injected diabetic rats (88). Compared to diabetic controls, rats given linalool showed a significant decrease in STZ-elevated plasma glucose and insulin levels, and a return of kidney carbohydrate-metabolizing enzymes and glycogen content to near normal levels. Moreover, in the kidney, linalool restored normal GLUT-1 expression, mitigated nephrin loss, suppressed oxidative stress and inflammation, and attenuated kidney damage and nephrotoxicity associated with STZ treatment. The authors suggested that linalool be considered as a potential therapy for renal damage in diabetics. Dyslipidemia is associated with diabetes and the metabolic syndrome. Oral administration of linalool (0.57 mg/d, 120 mg/d) for 6 weeks to mice fed a high-fat diet resulted in significantly lower plasma total and low-density lipoprotein cholesterol levels, compared to controls (89). Linalool also was evaluated for its hypotriglyceridemic action in mice fed a Western-diet (90). Compared to controls, mice administered linalool (100 mg/kg, p.o.) for 3 weeks showed a decrease in cumulative food intake, and a significant decrease in plasma triglyceride levels and saturated fatty acids. Additionally, compared to controls, linalool treatment significantly increased liver peroxisome proliferator-activated receptor (PPAR)a expression as well as that of several PPARa target genes. In in vitro assays, linalool was identified as a PPARa agonist through interaction with the PPARa ligand binding domain. The authors concluded that linalool can "rewire" the liver transcriptome and plasma metabolome. In a subsequent review (91) the authors proposed that linalool and other bioactive compounds can exert hypolipidemic and antiobesogenic effects by regulating PPARs.

References

1. Bajpal M, Mistra A, Prakash D. Antioxidant and free radical scavenging activities of some leafy vegetables. Int J Food Sci Nutr. 2005; 56: 473-481.

Wangensteen H, Samuelson A, Matterud K. Antioxidant activity in extracts from coriander.
 Food Chem. 2004; 88: 293-297.

3. Guera N, Melo E, Filho J. Antioxidant compounds from coriander (*Coriandrum sativum* L.) etheric extract. J Food Compos Anal. 2005; 18: 193-199.

 Dias M, Barros L, Sousa M, Ferreira I. Comparative study of lipophilic and hydrophilic antioxidants from *in vivo* and *in vitro* grown *Coriandrum sativum*. Plant Foods Food Hum Nutr.
 2011; 66: 181-186.

5. Baratta M,Dorman H, Deans S, Biondi D, Ruberto G. Chemical composition, antimicrobial and antioxidative activity of laurel, rosemary, oregano and cilantro essential oils. J Essent Oil Res. 1998; 10: 618-627.

6. Henning S, Zhang Y, Seeram N, Lee R, Wang P, Bowerman S, Heber D. Antioxidant capacity and phytochemical content of herbs and spices in dry, fresh, and blended herb paste form. Int J Food Sci Nutr. 2011; 62: 219-225.

7. Wong P, Kitts D. Studies on the dual antioxidant and antibacterial properties of parsley (*Petroselinum crispum*) and cilantro (*Coriandrum sativum*) extracts. Food Chem. 2006; 97: 505-515.

 Sharmin H, Nazma S, Mohiduzzaman N, Cadi P. Antioxidant capacity and total phenol content of commonly consumed selected vegetables of Bangladesh. Malays J Nutr.2011; 17: 377-383.

9. Satyanarayana S, Sushruta K, Sarma G, Srinivas N, Subba G. Antioxidant activity of the aqueous extracts of spicy food additives-evaluation and comparison with ascorbic acid in *in vitro* systems. J Herb Pharmacother. 2004; 4: 1-10.

 Tarwadi K, Agte V. Potential of commonly consumed green leafy vegetables for their antioxidant capacity and its linkage with the micronutrient profile. Int J Food Sci Nutr. 2003; 54: 417-425. 11. Ramadan M, Kroh L, Morsel J. Radical scavenging activity of black cumin (*Nigella sativa* L.) and niger (*Guizotia abyssinica* Cass.) crude seed oils and oil fractions. J Agric Food Chem.
2003; 51: 6961-6969.

12. Stashenko E, Puertas M, Martinez J. SPME determination of volatile aldehydes for evaluation of *in vitro* antioxidant activity. Anal Biochem Chem. 2002; 373: 70-74.

13. Park G, Kim H, Kim Y, Park S, Kim S, Oh M. *Coriandrum sativum* protects human keratinocytes from oxidative stress by regulating oxidative defense systems. Skin Pharmacol Physiol. 2012; 25: 93-99.

14. Hashim M, Lincy S, Remya V, Teena M, Anila L. Effect of polyphenolic compounds from *Coriandrum sativum* on H₂O₂-induced oxidative stress in human lymphocytes. Food Chem.
2005; 92: 653-660.

15. Reddy A, Lokesh B. Studies on spice principles as antioxidants in the inhibition of lipid peroxidation of rat liver microsomes. Mol Cell Biochem. 1992; 111:117-124.

 Krishnakantha T, Lokesh B. Scavenging of superoxide anions by spice principles. Indian J Biochem Biophys. 1993; 30: 133-134.

17. Usta J, Kreydiyyeh S, Knio K, Barnabe P, Bou-Moughlabay Y, Dagher S. Linalool decreases HepG2 viability by inhibiting mitochondrial complexes I and II, increasing reactive oxygen species and decreasing ATP and GSH levels. Chem Biol Interact. 2009; 180: 39-46.

18. Burt S. Essential oils: their antibacterial properties and potential applications in foods—a review. Int J Food Microbiol. 2004; 94: 223-253.

19. Reichling J, Schnitzler P, Suschke U, Saller R. Essential oils of aromatic plants with antibacterial, antifungal, antiviral, and cytotoxic properties – an overview. Forsch Komplementmed. 2009; 16: 79-90.

20. Deans S, Ritchie G. Antibacterial properties of plant essential oils. Int J Food Microbiol. 1987; 5: 165-180.

21. Delaquis P, Stanich K, Girard B, Mazza G. Antimicrobial activity of individual and mixed fractions of dill, cilantro, coriander, and eucalyptus essential oils. Int J Food Microbiol. 2002; 74: 101-109.

22. Chao S, Young D, Oberg C. Screening for inhibitory activity of essential oils on selected bacteria, fungi and viruses. J Essent Oil Res. 2000; 12: 639-649.

23. Matasyoh J, Maiyo Z, Ngure R, Chepkorir. Chemical composition and antimicrobial activity of the essential oil of *Coriandrum sativum*. Food Chem. 2009; 113: 526-529.

24. LoCantore P, Iacobellis N, DeMarco A, Capasso F, Senatore F. Antibacterial activity of *Coriandrum sativum* L. and *Foeniculum vulgare* Miller Var. vulgare (Miller) essential oils. J Agric Food Chem. 2004; 52: 7862-7866.

25. Elgayyar M, Draughon F, Golden D, Mount J. Antimicrobial activity of essential oils from plants against selected pathogenic and saprophytic microorganisms. J Food Prot. 2001; 64: 1019-1024.

26. Silva F, Ferreira S, Queiroz J, Domingues F. Coriander (*Coriandrum sativum* L.) essential oil: its antibacterial activity and mode of action evaluated by flow cytometry. J Med Microbiol. 2011; 60: 1479-1486.

27. Casetti F, Bartelke S, Biehler K, Augustin M, Schempp C, Frank U. Antimicrobial activity against bacteria with dermatological relevance and skin tolerance of the essential oil from *Coriandrum sativum* L. fruits. Phytother Res. 2012; 26: 420-424.

28. Duman A, Dayisoyluk K, Digrak M, Demirtas I, Alma M. Evaluation of bioactivity of linalool-rich essential oils from *Ocimum basilicum* and *Coriandrum sativum* varieties. Nat Prod Commun. 2010; 5: 969-974.

29. Kizil S, Sogut T. Investigation of antibacterial effects of some spices. Crop Res. 2003; 25:86-90.

30. Begnami A, Duarte M, Furletti V, Rehder V. Antimicrobial potential of *Coriandrum sativum*L. against different *Candida* species *in vitro*. Food Chem. 2010; 118: 74-77.

31. Park S, Lim Y, Freire M, Cho E, Jin D, Kook J. Antimicrobial effect of linalool and α -terpineol against periodontopathic and cariogenic bacteria. Anaerobe. 2012; 18: 369-372.

32. Carson C, Riley T. Antimicrobial activity of the major components of the essential oil of *Melaleuca acternifonlia*. J Appl Bacteriol. 1995; 78: 264-269.

33. Mukherjee K, Tribedi P, Mukhopadhyay B, Sil A. Antibacterial activity of long-chain fatty acids against mycobacteria. FEMS Microbiol Lett. 2013; 338: 177-183.

34. Bisignano G, Lagana M, Trombetta D, et al. *In vitro* antibacterial activity of some aliphatic aldehydes from *Olea europaea* L. FEMS Microbiol Lett. 2001; 198: 9-13.

59.

35. Kubo I, Fujita K, Kubo A, Nihei K, Ogura T. Antibacterial activity of coriander volatile compounds against *Salmonella choleraesuis*. J Agric Food Chem. 2004; 52: 3329-3332.

36. Duarte A, Ferreira S, Silva F, Domingues F. Synergistic activity of coriander oil and conventional antibiotics against *Acinetobacter baumanii*. Phyto medicine. 2012; 19: 236-238.
37. Chaudry N, Tariq P. Bactericidal activity of black pepper, bay leaf, aniseed and coriander against oral isolates. Pak J Pharm Sci. 2006; 19: 214-218.

38. Wong P, Kitts D. Studies on the antioxidant and antibacterial properties of parsley
(*Petroselinum crispum*) and cilantro (*Coriandrum sativum*) extracts. Food Chem. 2006; 97: 505515.

39. Rahman S, Parvez A, Islam R, Khan M. Antibacterial activity of natural spices on multiple drug resistant *Escherichia coli* isolated from drinking water-Bangladesh. Ann Clin Microbiol Antimicrob. 2011; 10:10.

40. O'Mahony R, Al-Khtheeri H, Weerasekera D, et al. Bactericidal and anti-adhesive properties of culinary and medicinal plants against *Helicobacter pylori*. World J Gastroenterol. 2005; 11: 7499-7507.

41. Saeed S, Tariq P. Antibacterial activities of *Emblica officinalis* and *Coriandrum sativum* against gram negative urinary pathogens. Pak J Pharm Sci. 2007; 20: 32-35.

42. Kim Y, Kang S, Choi O. Antimicrobial activity of coriander (*Coriandrum sativum* L.) extract. J Kor Soc Food Nutr. 2001; 30: 692-696.

43. Silva F, Ferreira S, Duarte A, Mendonca D, Domingues F. Antifungal activity of *Coriandrum sativum* essential oil, its mode of action against *Candida* species and potential synergism with amphotericin B. Phytomedicine. 2011; 19: 42-47.

44. Minija J, Thoppil J. Volatile oil constitution and microbiological activities of essential oils of *Coriandrum sativum* L. J Natur Remed. 2001; ¹/₂: 147-150.

45. Hsu C, Lai W, Chuang K, Lee M, Tsai Y. The inhibitory activity of linalool against
filamentous growth and biofilm formation in *Candida albicans*. Med Mycol. 2012; in press.
46. Furletti V, Teixeira I, Obando-Pereda G, Mardegan R, et al. Action of *Coriandrum sativum*L. essential oil upon oral *Candida albicans* biofilm formation. Evid Based Complement Altern
Med. 2011; doi:10.1155/2011/985832.

47. Martin K, Ernst E. Antiviral agents from plants and herbs: a systematic review. Antivir Ther. 2003; 8:77-90.

48. Rondon F, Bevilaqua C, Accioly M, Morais S, et al. *In vitro* effect of Aloe vera, *Coriandrum sativum* and *Ricinus communis* fractions on *Leishmania infantum* and on murine monocytic cells. Vet Parasitol. 2011; 178: 235-240.

49. Fernandez-Pan I, Mate J. Antimicrobial activity of whey protein isolate edible films with essential oils against food spoilers and foodborne pathogens. J Food Sci. 2012; 77: M383-M387.
50. Rattanachaikunsopon P, Phumkhachorn P. Potential coriander (*Coriander sativum*) oil as a natural antimicrobial compound in controlling *Campylobacter jejuni* in raw meat. Biosci Biotechnol Biochem. 2010; 74: 31-35.

51. Al-Jedah J, Ali M, Robinson R. The inhibitory action of spices against pathogens that might be capable of growth in a fish sauce (mehiawah) from the Middle East. Int J Food Microbiol. 2000; 57: 129-133.

52. Conner D, Beuchat L. Effects of essential oils from plants on growth of food spoilage yeast. J Food Sci. 1984; 49: 429-434.

53. Gill A, Delaquis P, Russo P, Holley R. Evaluation of antilisterial action of cilantro oil on vacuum packed ham. Int J Food Microbiol. 2002; 73: 83-92.54e.

54. Klein G, Ruben C, Upmann M. Antimicrobial activity of essential oil components against potential food spoilage microorganisms. Curr Microbiol. 2013; doi 10.1007/s00284-013-0354-1.
55. Michalczyk M, Macura R, Tesarowicz I, Banas J. Effect of adding essential oils of coriander (*Coriandrum sativum* L.) and hyssop (*Hyssopus officinalis* L.) on the shelf life of ground beef. Meat Sci. 2012; 90: 842-850.

56. Lixandru B, DraceaN, Dragomirescu C, Dragulescu E, et al. Antimicrobial activity of plant essential oils against bacterial and fungal species involved in food poisoning and/or food decay. Roum Arch Microbiol Immunol. 2010; 69: 224-230.

57. Yin M, Cheng W. Inhibition of *Aspergillus niger* and *Aspergillus flavus* by some herbs and spices. J Food Prot. 1998; 61: 123-125.

 58. Basilico M, Basilico J. Inhibitory effects of some spice essential oils on *Aspergillus* ochraceus NRRL 3174 growth and ochratoxin A production. Lett Appl Microbiol. 1999; 29: 238-241.

59. Coelho V, Mazzardo-Martins L, Martins D, et al. Neurobehavioral and genotoxic evaluation of (-)-linalool in mice. J Nat Med. 2013; DOI 10.1007/s11418-013-0751-6

60. Guzman-Gutierrez S, Gomez-Cansino R, Garcia-Zebadua J, et al. Antidepressant activity of *Litsea glaucescens* essential oil: identification of β -pinene and linalool as active principles. J Ethnopharmacol. 2012; 143: 673-679.

61. Cline M, Taylor J, Flores J, et al. Investigation of the anxiolytic effects of linalool, a lavender extract, in the male Sprague-Dawley rat. AANA J. 2008; 76: 47-51.

62. Kaewwongse M, Sanesuwan K, Pupa P, Bullangpoti V. Essential oil compounds as stressreducing agents in rats. Comm Appl Biol Sci Ghent Univ. 2013; 78: 167-172.

63. Linck V, DaSilva A, Figueiro M, et al. Inhaled linalool-induced sedation in mice.

Phytomedicine. 2009; 16: 303-307.

64. Tankam J, Ito M. Inhalation of the essential oil of *Piper guineense* from Cameroon shows sedative and anxiolytic-like effects on mice. Biol Pharm Bull. 2013; 36: 1608-1614.

65. Buchbauer G, Jirovetz L, Jager W, et al. Aromatherapy: evidence for sedative effects of the essential oil of lavender after inhalation. Z Naturforsch. 1991; 46: 1067-1072.

66. Linck V, DaSilva A, Figueiro M, et al. Effects of inhaled linalool on anxiety, social interaction, and aggressive behavior in mice. Phytomedicine. 2010; 17: 679-683.

67. Coelho V, Gianesini J, VonBorowski R, et al. (-)-Linalool, a naturally occurring monoterpene compound, impairs memory acquisition in the onject recognition test, inhibitory avoidance test and habituation to a novel environment in rats. Phytomedicine. 2011; 18: 896-901.
68. Peana A, D'Aquila P, Chessa M, et al. (-)-Linalool produces antinociception in two experimental models of pain. Eur J Pharmacol. 2003; 460: 37-41.

69. Peana A, DeMontis M, Nieddu E, et al. Profile of spinal and supraspinal antinociception of (-)-linalool. Eur J Pharmacol. 2004; 486: 165-174.

70. Peana A, Rubattu P, Piga G, et al. Involvement of A1 and A2A receptors in (-)-linaloolinduced antinociception. Life Sci. 2006; 78: 2471-2474.

71. Quintans-Junior L, Barreto R, Menezes P, et al. β-cyclodextrin-complexed (-)-linalool produces antinociceptive effect superior to that of (-)-linalool in experimental pain protocols. Basic Clin Pharmacol Toxicol. 2013; 113: 167-172.

72. Nascimento S, Camargo E, DeSantana J, et al. Linalool and linalool complexed in β cyclodextrin produce anti-hyperalgesic activity and increase Fos protein expression in animal model for fibromyalgia. Naun Schmied Arch Pharmacol. 2014; 387: 935-942.

73. Guzman-Gutierrez S, Bonilla-Jaime H, Gomez-Cansino R, Reyes-Chilpa R. Linalool and β-pinene exert their antidepressant-like activity through the monaminergic pathway. Life Sci.
2015; 128: 24-29.

74. Re L, Barocci S, Sonnino S, et al. Linalool modifies the nicotinic receptor-ion channel kinetics at the mouse neuromuscular junction. Pharmacol Res. 2000; 42: 177-182

75. Yamada K, Mimaki Y, Sashida Y. Effects of inhaling the vapor of *Lavandula burnatii super*derived essential oil and linalool on plasma adrenocorticotropic hormone (ACTH), catecholamine and gonadotrophin levels in experimental menopausal female rats. Biol Pharm Bull. 2005; 28: 378-379.

76. Nakamura A, Fujiwara S, Marsumoto I, Abe K. Stress repression in restrained rats by R-(-)linalool inhalation and gene expression profiling of their whole blood cells. J Agric Food Chem. 2009; 57: 5480-5485.

77. Nakamura A, Fujiwara S, Ishijima T, Okada S, et al. Neuron differentiation-related genes are up-regulated in the hypothalamus of odorant-inhaling rats subjected to acute restraint stress. J Agric Food Chem. 2010; 58: 7922-7929.

78. Yamamoto N, Fujiwara S, Saito-Iizumi K, et al. Effects of inhaled (S)-linalool on
hypothalamic gene expression in rats under restraint stress. Biosci Biotechnol Biochem. 2013;
77: 2413-2418.

79. Batista P, Werner M, Oliviera E, et al. Evidence for the involvement of iontropic glutamate receptors on the antinociceptive effect of (-)-linalool in mice. Neurosci Lett. 2008; 440: 299-303.

80. Sakurada T, Mizoguchi H, Kuwahata H, et al. Intraplantar injection of bergamot essential oil induces peripheral antinociception mediated by opoid mechanism. Pharmacol Biochem Behav. 2011; 97: 436-443.

81. Katsuyama S, Otowa A, Kamio S, et al. Effect of plantar subcutaneous administration of bergamot essential oil and linalool on formalin-induced nociceptive behavior in mice. Biomed Res. 2105; 36: 47-54.

82. Batista P, Werner M, Oliviera E, et al. The antinociceptive effect of (-)-linalool in models of chronic inflammatory and neuropathic hypersensitivity in mice. J Pain. 2010; 11: 1222-1229.

83. Peana A, DeMontis M, Sechi S, et al. Effects of (-)-linalool in the acute hyperplasia induced by carrageenan, L-glutamate and prostaglandin E₂. Eur J Pharmacol. 2004; 497: 279-284.

84. Katsuyama S, Kuwahata H, Yagi T, et al. Intraplantar injection of linalool reduces paclitaxelinduced acute pain in mice. Biomed Res. 2012; 33: 175-181.

85. Kuwahata H, Komatsu T, Katsuyama S, et al. Peripherally injected linalool and bergamot essential oil attenuate mechanical allodynia via inhibiting spinal ERK phosphorylation. Pharmacol Biochem Behav. 2013; 103: 735-741.

86. Berliocchi L, Russo R, Levato A, et al. (-)-Linalool attenuates allodynia in neuropathic pain induced by spinal nerve ligation in C57/BL6 mice. Int Rev Neurobiol. 2009;

DOI:10.1016/S0074-7742(09)85017-4

87. Peana A, D'Aquila P, Panin F, et al. Anti-inflammatory activity of linalool and linalyl acetate constituents of essential oils. Phytomedicine. 2002; 9: 721-726.

 Deepa B, Anuradha C. Effects of linalool on inflammation, matrix accumulation and podocyte loss in kidney of streptozotocin-induced diabetic rats. Toxicol Mech Meth. 2013; 23: 223-234.

89. Cho S, Jun H, Lee J, et al. Linalool reduces the expression of 3-hydroxy-3-methylglutaryl CoA reductase via sterol regulatory element binding protein-2- and ubiquitin-dependent mechanisms. FEBS Lett. 2011; 585: 3289-3296.

90. Jun H, Lee J, Kim J, et al. Linalool is a PPARα ligand that reduces plasma TG levels and rewires the hepatic transcriptome and plasma metabolome. J Lipid Res. 2014; 55: 1098-1110.

91. Lee S, Jia Y. The effect of bioactive compounds in tea on lipid metabolism and obesity through regulation of peroxisome proliferator-activated receptors. Curr Opin Lipidol. 2015; 26: 3-9.