

Supplemental Material

Methods:

Determination of *S. aureus* load in tissues by quantitative PCR (qPCR):

For real-time PCR detection, log phase culture of *S. aureus* (10^9 CFU) was serially diluted and DNA was isolated by previously described protocol with minor modifications [1]. Briefly, bacterial cells were harvested lysed in 100 μ l of NET buffer (50 mM NaCl, 125 mM EDTA, 50 mM Tris-HCl [pH 7.6]) containing 50 μ l of 2.6 N NaOH solution and 100 μ l of 24% sodium dodecyl sulphate (SDS; final concentration 3.4%). Lysates were incubated at 80°C for 10 min. and cooled to room temperature before adding proteinase K (200 μ g/ ml) for digestion (50°C for 1.5 hour). DNA extracted from the lysates by standard phenol-chloroform, precipitated with isopropanol was quantified at A260 for analysis by real-time PCR to generate a standard curve (Bacterial CFU versus Ct value; Supplemental figure 1). To determine the *S. aureus* load in tissues, bacterial DNA isolated from the infected host tissue samples using the above protocol were subjected to real-time PCR. The Ct values thus obtained were used to determine the CFU in infected tissues from the standard curve generated as described above. For detection of *S. aureus* strain in brain, different parts of brain tissues like hippocampus, prefrontal and cerebral cortex, cerebellum, brain stem, parts of midbrain were mixed and homogenised.

Extracted *S. aureus* DNA was subjected to real-time PCR using primer that targets Staphylococcus 16S rRNA gene STPY [2]. The sequences of the primers used are provided below:

STPYF: 5'-ACGGTCTTGCTGTCACCTTATA-3' and

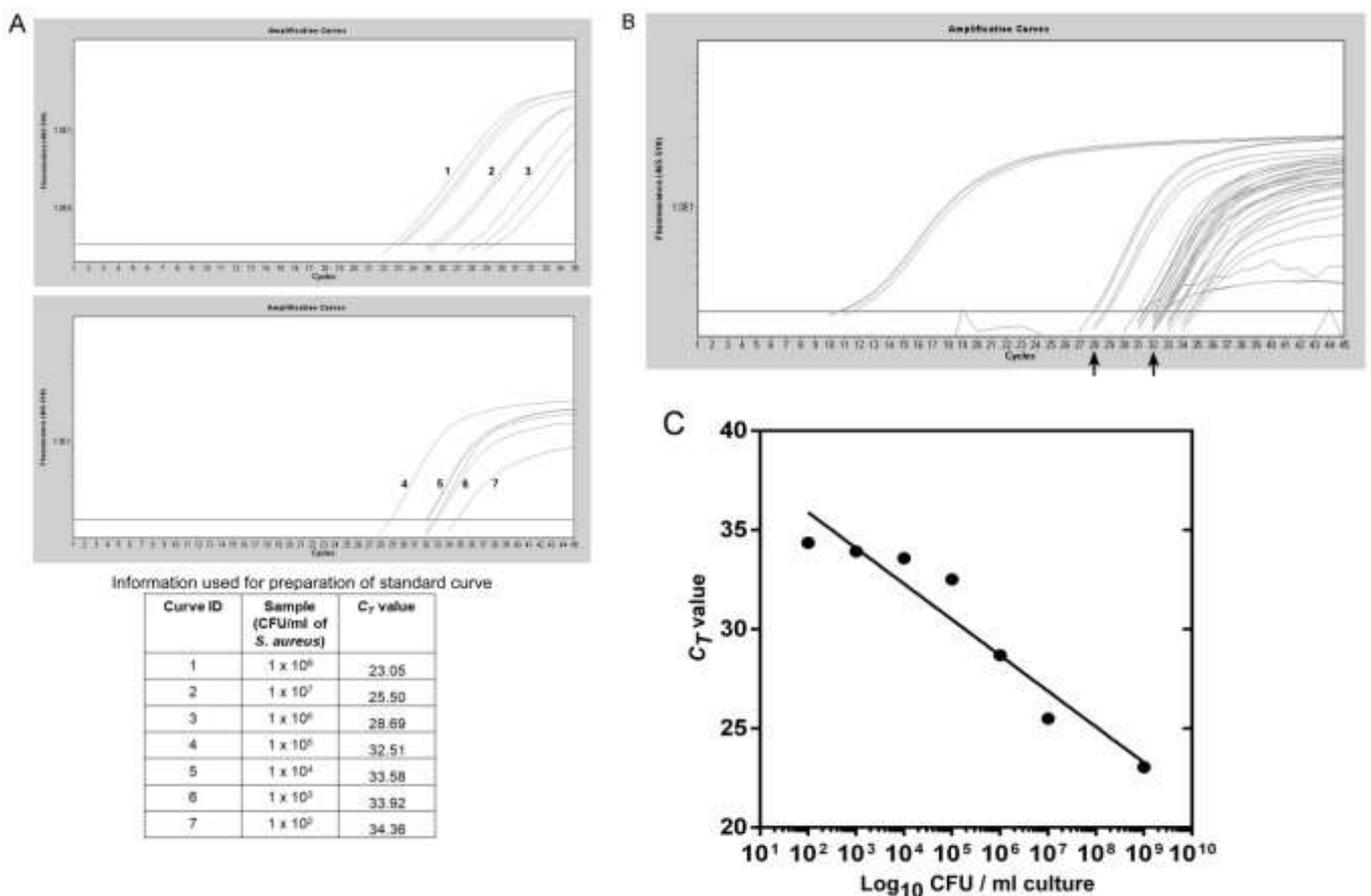
STPYR2: 5'- TACACATATGTTCTTCCCTAATAA-3'

Real-time PCR was performed using 2X SYBR green master mix (KAPA SYBR FAST kit, KAPA BIOSYSTEMS, USA) in Light Cycler® 480 real-time PCR machine (Roche, Germany) with 1 μ l of DNA samples in 10 μ l reaction volume for 35 cycles of amplification [denaturation at 95°C for 10 sec; annealing at 57°C for 20 sec; extension at 72°C for 2 sec].

References:

- [1] Romero C, Lopez-Goñi I. Improved method for purification of bacterial DNA from bovine milk for detection of *Brucella* spp. by PCR. *Appl Environ Microbiol.* 1999; 65:3735-7
- [2] Johnson EJ, Zemanick ET, Accurso FJ, Wagner BD, Robertson CE, Harris JK. Molecular identification of *Staphylococcus aureus* in airway samples from children with cystic fibrosis. *PLoS One* 2016; 11:1–12. doi:10.1371/journal.pone.0147643.

Supplementary Figure 1: Real time PCR amplification curve and standard curve for determination of bacterial load in tissues. (A) Amplification curves of real-time PCR generated for determination of C_T values for *S. aureus* STPY gene. Genomic DNA isolated from serially diluted log phase culture (10^9 CFU) of *S. aureus* were used for amplification using STPY gene specific primers. Intercept of the magnitude of the fluorescence signal with the horizontal threshold line represents the C_T value.



Number indicated in two graphs were curve ID of samples (10^9 CFU- 10^2 CFU *S. aureus*), used for plotting the standard curve. All the corresponding C_T values were listed in the table. (B) Amplification

curve of real-time PCR showing corresponding C_T values for STPY gene products in spleen and blood samples indicated by arrows. (C) Standard curve for determination of bacterial load in tissues was generated by plotting C_T values from real-time PCR vs CFU per ml of culture. Each point represents the amount of DNA present in a given CFU of *S. aureus* corresponding to the C_T value. CFU = Colony Forming Unit.

Supplementary Figure 2

Histone acetylation and tyrosine hydroxylase expression profile in hippocampus of control animals with spectinomycin [Spec (+)] versus without spectinomycin [Spec (-)]. (A) Specific acetylated residues (K9, K27 and K14) of histone H3 were examined separately on immunoblots to quantify the residue-specific acetylation with respect to the total histone H3. Data presented as mean \pm SEM. (B) Tyrosine hydroxylase (TH) protein expression in control animals with Spec vs without Spec. TH band intensities were normalized by GAPDH signal. (n = 4). Data was presented as mean \pm SEM. Spec = Spectinomycin.

